Gas sensing using polymer-functionalized deformable Fabry–Perot interferometers

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ARTICLE INFO

Article history:

Received 1 October 2012

Received in revised form

31 December 2012

Accepted 4 February 2013

Available online xxx

Keywords:

Gas sensing

Fabry–Perot interferometer

Polymer swelling

Volatile organic compound

Silicon micromachining

ABSTRACT

We report a chemical vapor sensor in which polymer swelling, upon analyte absorption, is used to deform an on-chip silicon Fabry–Perot interferometer (FPI). The magnitude of the deformation, recorded through the resonance wavelength shift, is proportional to the analyte concentration in accordance with a simplified analytical model and with finite element simulations. Conventional and phenyl-doped polydimethylsiloxane (PDMS) polymers are used to functionalize different interferometers, which are tested for the detection of two volatile organic compounds, i.e. m-xylene and cyclohexane. The detection of m-xylene concentrations down to 34 ppm—limited by our flow-meter setup—is achieved experimentally. Based on the sensitivities and the noise characteristics of the devices, limits of detection (LODs) of 1.6 ppm m-xylene and 6.3 ppm cyclohexane are expected.

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1. Introduction

Upon analyte absorption, many polymers (e.g., PDMS) are known to expand [1], a phenomenon usually referred to as “polymer swelling”. Such expansion can be used to deform optical interferometers, causing a shift of their resonance condition that can be monitored to create chemical sensors. This approach was previously used with various optical interferometers. Polymer-coated fiber Bragg gratings were shown to deform upon exposition to salt [2] hydrocarbons [3] and humidity [4]. Bragg gratings were also fabricated directly with deformable polymer layers and were found to be sensitive to the presence of acetone [5], salt [6], and volatile organic compounds [7] (VOCs). Fabry–Perot interferometers (FPI) can also be formed simply by exploiting reflections at the two material interfaces of single deformable polymer membranes. This approach was used to detect VOCs [8–10] and ionic strength [11].

The sensors reported here rely on deformable Fabry–Perot interferometers. As presented in Fig. 1, these FPIs are implemented in-plane with the substrate by vertical plasma etching of silicon-air Bragg reflectors. The in-plane (i.e.: vertical mirrors, in-plane

optical axis) rather than out-of-plane [5–11] configuration allows monolithic integration of microfluidic systems and optical fiber alignment grooves. Integrated microfluidic systems simplify the parallel functionalization of multiple interferometers, using different polymers, to target specific classes of analytes. Fiber grooves allow the remote interrogation of the interferometers with passively aligned conventional single-mode optical fibers, rather than requiring free space alignment of light sources and photodetectors [5–11].

2. Methodology

2.1. Sensing mechanism

Upon analyte absorption, three factors can cause shifts of the interferometer ($\Delta\lambda_{\text{Res}}$) resonance wavelength. First, as depicted in Fig. 1(b), polymer swelling deforms the interferometer, resulting in an increase (ΔL) of the mirror separation (L). This mechanism will later be demonstrated to be the dominant one. Secondly, if the refractive index of the analyte (n_a) and the polymer (n_p) are different, the refractive index of the polymer–analyte mixture (n) should change by a factor Δn_{Mix} upon sample absorption. Lastly, if the mirrors oppose a significant force to the polymer expansion, there should be a densification of the polymer, which should lead to a Δn_{σ} variation of its refractive index. All three effects change

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Fig. 1. (a) Schematic representation of the deformable interferometric sensor. (b) Schematic representation of the sensor operation principle. Upon sample absorption, polymer expansion deforms the interferometer, inducing a shift of its resonance wavelength. Refractive index changes in the polymer, upon absorption, also contribute to this shift.

the interferometer optical pathlength (nL), and contribute to the measured resonance wavelength shift ($\Delta\lambda_{\text{Res}}$) through:

$$\frac{\Delta\lambda_{\text{Res}}}{\lambda_{\text{Res}}} = \alpha \frac{\Delta L}{L} + \beta \frac{(\Delta n_{\text{Mix}} + \Delta n_{\sigma})}{n}, \quad (1)$$

where $\alpha=0.75$ and $\beta=0.93$ are device-specific reduction factors (≤ 1). The factor α accounts for the fact that only one layer in each Bragg mirror is displaced upon analyte absorption (see Fig. 1b). The factor β accounts for the fact that the resonant mode inside the cavity is not located entirely inside the polymer, but also partly inside the mirrors (i.e., the thickness of the mirrors is not negligible compared to L , the distance between the mirrors). The value of both parameters can be calculated using a previously reported optical simulation algorithm for deep-etched Fabry–Perot interferometers [12], as detailed in Appendix A1.

We define the dimensionless sensitivity (Γ_{ϕ}) as the normalized ratio of the resonance wavelength shift ($\Delta\lambda_{\text{Res}}$) to the absorbed volume fraction of analyte (ϕ_a) inside the polymer:

$$\Gamma_{\phi} = \frac{1}{\lambda_{\text{Res}}} \frac{\Delta\lambda_{\text{Res}}}{\Delta\phi_a} \quad (2)$$

Conveniently, ϕ_a can be related to the volume/volume (v/v) concentration of analyte in air (C_{air}), near the polymer, using the ideal gas law and the partition coefficient of the polymer for the respective analyte (K_{p-a}):

$$\phi_a = K_{p-a} \frac{PV_m}{RT} C_{\text{air}} \quad (3)$$

where P is the pressure (1 atm in our case), T is the temperature, R is the gas constant, and V_m is the molar volume of the analyte, in liquid phase.

For all the calculations presented in this work we assume that upon absorption, the final volume of the polymer–analyte mixture equals the initial volume of the polymer, plus the absorbed volume of analyte. In other words we assume that there is no volume reduction upon absorption, except in the presence of mechanical stresses inside the polymer (calculations in Section 2.1.1 will demonstrate these stresses lead to negligible volume reductions). In this context, the relative volume (V) expansion of the polymer upon absorption of a ϕ_a volume fraction of analyte is given by:

$$\frac{\Delta V}{V} = \Delta\phi_a \quad (4)$$

This assumption of volume additivity is supported by reports that the absorption of a slightly lower refractive index analyte

(cyclohexane) reduced the refractive index of a PDMS-based polymer [13]. This would not have been possible if significant volume reduction occurred upon absorption, since volume reduction would have led to an increase of the refractive index.

2.1.1. Simplified analytical model

This section details the sensitivity (Γ_{ϕ}) of deformable FPI sensors in the case where:

- (1) the mirrors are perfectly movable, such that they oppose negligible forces to the expansion of the polymer,
- (2) the volume increase of the polymer (i.e. swelling) is directed mainly in one direction, parallel to the optical pathlength between the mirrors (L), such that ΔL is maximized.

Interestingly, these conditions straightforwardly apply to out-of-plane deformable Fabry–Perot interferometers [8–11]. In these cases, the polymer is bonded to a rigid substrate such that swelling can only occur in the out-of-plane direction, which is parallel to the optical axis (criterion 2). There is also no force opposing the movement of the top material interface that is used as the movable mirror of the interferometer (criterion 1).

For the in-plane devices presented in the current work (Fig. 1), compliance with the two above mentioned criteria is less trivial. The deformable layer in each Bragg mirror might oppose non-negligible force to the expansion of the polymer. The polymer might also swell upward (positive z direction in Fig. 1a) if the bonding strength with silicon is not strong enough. We will therefore use the simplified ideal formalism presented in this section primarily as a basis for comparison of our in-plane sensor with out-of-plane sensing mechanisms. In Section 2.1.2 finite element simulations will address the limitations of the analytical model for in-plane devices.

Under the two conditions listed above, it is possible to show that the relative length increase between the mirrors, upon absorption of a ϕ_a volume fraction of analyte, is given by:

$$\left. \frac{\Delta L}{L} \right|_{\text{Simplified}} = \frac{\Delta\phi_a}{3} \frac{1+\nu}{1-\nu} \quad (5)$$

where ν is the polymer Poisson's ratio. It is also possible to show that the compressive stress experienced by the polymer due to its

restrained expansion in the directions perpendicular to the optical axis (y and z in Fig. 1) is given by:

$$\sigma_{\perp} = \frac{\phi_a E}{3(1-\nu)} \quad (6)$$

where E is the Young's modulus of the polymer. This stress will in turn lead to a volume reduction of the polymer given by:

$$\frac{\Delta V_h}{V} = \frac{\Delta \sigma_h}{B} = \frac{2}{9} \frac{\Delta \phi_a E}{B(1-\nu)} \quad (7)$$

where B is the bulk modulus of the polymer, and $\sigma_h = 2/3 \sigma_{\perp}$ is the hydrostatic stress inside the polymer.

Also, the addition of a ϕ_a analyte volume fraction will lead to a Δn variation of the polymer refractive index. Several rules exist to predict the refractive index of binary mixtures [14,15]. The Lorentz–Lorenz relation is probably most frequently used. We however choose to use the simpler Gladstone–Dale relation, which yields very similar results for the typical refractive indices that we will encounter. This relation (Eq. (8)) is linear relative to ϕ_a , leading to a simplified expression of the sensitivity (Γ_{ϕ}).

$$\frac{\Delta n_{\text{Mix}} + \Delta n_{\sigma}}{n} = \Delta \phi_a \frac{(n_a - n_p)}{n_p} + \frac{\Delta V_h}{V} \frac{(n_p - 1)}{n_p} \quad (8)$$

Finally, combining Eqs. (1, 5–8) in Eq. (2) yields the sensitivity of deformable Fabry–Perot chemical sensors in the context of the simplified analytical model:

$$\Gamma_{\phi}|_{\text{Simplified}} = \frac{\alpha}{3} \frac{1+\nu}{1-\nu} + \beta \frac{(n_a - n_p)}{n_p} + \beta \frac{2}{9} \frac{(n_p - 1)}{n_p} \frac{E/B}{1-\nu} \quad (9a)$$

In Eq. (9a), the first term accounts for interferometer deformations. The second term accounts for polymer refractive index variations due to mixture with an analyte of different refractive index. The third term accounts for refractive index variations due to hydrostatic stress upon expansion.

The absorbent polymers used in the following experiments are essentially incompressible. For example, the bulk modulus [16] ($B = 10^3$ MPa) of Sylgard 184 PDMS is significantly higher than its Young's modulus [17] ($E = 1.8$ MPa), which yield a $\nu = 0.5 - E/2B = 0.499$ Poisson's ratio. In this context, the third term of Eq. (9a) is almost 3 orders of magnitude smaller than the first two terms and can be neglected. Eq. (9a) can therefore be rewritten as:

$$\Gamma_{\phi}|_{\text{Simplified}} \approx \alpha + \beta \frac{(n_a - n_p)}{n_p} \quad (9b)$$

Furthermore, for most organic compounds, the refractive index difference in Eq. (9b) ($n_a - n_p$) should be in the order of 0–0.15. In these cases, the term that accounts for interferometer deformations (α) should always be at least 10 times larger than the term that accounts for refractive index variations. Therefore, with $\alpha = 0.75$, the absorption of, for example, a $\phi_a = 1\%$ volume fraction of analyte is expected to induce a $\Delta \lambda_{\text{res}}/\lambda_{\text{res}} \approx 0.75\%$ shift of the interferometer resonance wavelength.

2.1.2. Finite element simulations

In order to evaluate if the response of the fabricated devices can be expected to be close to the simplified analytical case, finite element simulations are performed using CoventorWare 2010. An isotropic expansion of the polymer volume filling the gap between the two mirrors (see Fig. 1) is simulated. Upon expansion, the relative increase of the distance between the mirrors ($\Delta L/L$) and the hydrostatic pressure inside the polymer (σ_h) are evaluated. For these simulations, we use the Young's modulus ($E = 1.8$ MPa) [17] and Poisson's ratio ($\nu = 0.499$) values of Sylgard 184 PDMS.

The meshed model used for the simulations is presented in Fig. 2(a). For each of the two Bragg mirrors, only the one silicon layer that is in contact with the polymer is included in the model.

The (y, z) dimensions of these layers are ($130 \mu\text{m}, 80 \mu\text{m}$). Their thickness (in the x direction) is 600 nm , as determined by previous optical characterization of the interferometers [12]. The layers are attached at both y extremities to the edges of the microfluidic channel, which are considered to be immobile. The bottom z extremities of the mirror layers are fixed to the substrate (Fix All boundaries in Fig. 2a) while their upper z extremities are free to move.

Two different types of boundary conditions are used to simulate the contact interfaces between silicon and PDMS. The first type considers the two materials to be tightly bonded together, such that no sliding can occur. The second type considers the two materials to be in contact but to be free to slide on each other without friction. We expect the experimental results to lie somewhere between these two extreme cases, which we will refer to as the “bonded” and “sliding” models.

Only a small portion of the microfluidic channel length ($40 \mu\text{m}$) is included in the mesh model on each side of the Fabry–Perot cavity. The continuation of the truncated channel must therefore be simulated using appropriate boundary conditions. It is possible to show that, away from the deformable mirror layers, no polymer movement occurs in the directions (x, y) parallel to the substrate due to the rigid boundaries of the microfluidic system. The continuation of the channels is therefore simulated using “Fix Y” boundary conditions (see Fig. 2a).

For the two models, the response to various polymer volume increases is simulated. A linear relation is established between the absorbed volume fraction of analyte (ϕ_a) and the relative path-length increase between the mirrors (on the optical axis of the system):

$$\begin{aligned} \left. \frac{\Delta L}{L} \right|_{\text{Bonded}} &= 1.77 \Delta \phi_a, \\ \left. \frac{\Delta L}{L} \right|_{\text{Sliding}} &= 0.51 \Delta \phi_a. \end{aligned} \quad (10)$$

A linear relation is also established between ϕ_a and the hydrostatic stress (σ_h) inside the polymer volume where light will interact with the system (around the optical axis in Fig. 2a). As for the simplified analytical model, the hydrostatic stress (σ_h) is found to be negligible compared with the bulk modulus of PDMS based polymers. Its contribution to the sensitivity of the sensors can therefore still be neglected, as in Section 2.1.1.

Finally, replacing Eq. (5) by Eq. (10) in the formalism of Section 2.1.1 yields the sensitivity of the sensors (Γ_{ϕ}) for both the bonded and the sliding models:

$$\begin{aligned} \Gamma_{\phi}|_{\text{Bonded}} &= 1.77\alpha + \beta \frac{(n_a - n_p)}{n_p}, \\ \Gamma_{\phi}|_{\text{Sliding}} &= 0.51\alpha + \beta \frac{(n_a - n_p)}{n_p}. \end{aligned} \quad (11)$$

In Eq. (11) the sliding model yields a deformation sensitivity (0.51α) that is twice smaller than what was predicted (1.00α) by the simplified analytical model. This result was expected since the polymer is now free to expand in two directions (x and z) rather than only one for the analytical model (similarly, a polymer that would be free to expand in all three dimensions would yield a 0.33α sensitivity).

Surprisingly, the bonded model yields a higher deformation sensitivity (1.77α) than what is predicted by the simplified analytical model in Eqs. (9a) and (9b) (1.00α). This is because, in Fig. 2(b), the polymer is not free to expand in the microchannel, on each side of the optical cavity. Consequently, the polymer located in these regions expands towards the optical axis of the system, where the deformation is amplified (see Appendix A2). However, this behavior is found to depend strongly on the polymer Poisson's ratio. For $\nu < 0.499$, the predicted sensitivity of the bonded model quickly

Fig. 2. (a) Mesh model and boundary conditions for the simulations of the interferometer deformation upon polymer swelling. (b and c) Magnitude of the deformation upon a 2% polymer volume increase. The deformations (but not the color scales) are exaggerated by a factor 5. The simulations are performed for bonded (b) and sliding (c) silicon-PDMS material interfaces.

reduces and essentially matches the sensitivity of the analytical model when $\nu < 0.45$ (see Appendix A2).

2.2. Experimental methodology

A scanning electron microscope (SEM) photograph of a Fabry–Perot interferometer is presented in Fig. 3(b). This device has 3 silicon layers per mirror, compared to only two in the schematic view of Fig. 1. Both configurations were fabricated and tested for gas detection and, as predicted in Appendix A1, were found to respond similarly to deformations and refractive index changes. The interferometers are etched in silicon wafers using an inductively coupled plasma reactor (ICP180-100, Oxford Instruments Inc.). A two-mask soft lithography process is used to fabricate successively the smaller (multilayer mirrors) and the larger (optical fiber alignment grooves, microfluidic system) features [12].

The absorbent polymer (Fig. 3(c)) is inserted between the two mirrors of the interferometer using a monolithically integrated microfluidic system. A drop ($\sim 15\text{--}20\ \mu\text{L}$) of liquid pre-polymer mixture is placed in the leftmost reservoir of Fig. 3(a). The polymer flows by capillary forces in the microfluidic channel to fill the gap in the interferometer (Fig. 3(c)), and is then left to cure at room temperature for several hours. We did not determine the minimum required polymer curing time. Some devices required more than 12 h of curing for stable performance, so all devices were cured for more than a week.

Two different polymers were tested for the functionalization of the interferometers. Some devices were coated with commercial PDMS (Dow Corning® Sylgard 184), while the others were functionalized with a PDMS–polydiphenylsiloxane copolymer (PDMS–PDPS). The copolymer was prepared by a procedure similar to the one previously reported [13], but using different mixture proportions (10% polydiphenylsiloxane (PDPS) mole fraction, 3% titanium tetraisopropoxide cross-linker mole fraction).

The functionalized devices were tested for the detection of two different vapor phase analytes at room temperature and

atmospheric pressure. To produce different analyte concentrations, saturated vapor (10,720 ppm for m-xylene, 125,340 ppm for cyclohexane [13]) was first generated by bubbling nitrogen through a wash bottle containing the analyte in liquid phase. A two-channel mixing flowmeter (Praxair PRXFM-4621(22)) was then used to dilute back the saturated vapor with pure nitrogen, in order to lower analyte concentration. The first channel of the flowmeter (nitrogen) uses a tantalum bead float, while the second channel (saturated analyte vapor) uses a glass float. The flowmeter and gas bubbling assembly are shown in Fig. 4(a). A gas cap (Fig. 4(b)) was lowered over the sensor to allow analyte vapor to disperse evenly over the chip. The complete setup was housed inside a laminar flow hood. Cyclohexane and m-xylene were chosen as the analytes to allow comparison with previous work using similar polymers [13,18].

Upon gas exposure, the interferometer resonance wavelength was tracked in real time by continuously scanning a tunable external-cavity diode laser source (Ando AQ4320D). The laser was interfaced with the interferometer through conventional single-mode optical fibers (Corning® SMF-28), which were cleaved and inserted in the optical fiber alignment grooves (Fig. 3). The transmission spectrum was recorded at 30 s intervals using an InGaAs photodiode detector (Thorlabs D400FC) and a lock-in amplifier (Stanford Research Systems SRS844 RFCA). The voltage output of the lock-in amplifier was then collected using a 16-bit data acquisition USB device (Measurements Computing PMD 1608 FS) and custom-made LabView (National Instruments) software. Amplitude noise was minimized numerically in each recorded spectrum using a zero-phase shift, forward and reverse digital convolution filter (Matlab® “FILTfilt” function). A 1 nm wide Gaussian distribution ($\sigma = 0.2\ \text{nm}$) was used as the filter. The exact resonance wavelength position was then determined for each spectrum by fitting a 4th order polynomial to the top half of the filtered resonance peaks. Results and discussion

The response of a PDMS-coated interferometer to various m-xylene and cyclohexane vapor concentrations is presented in Fig. 5.

Fig. 3. (a) Low magnification scanning electron micrograph of a sensor, prior to polymer coating, showing the monolithically integrated microfluidic system used for functionalization. (b) Higher magnification view of the Fabry–Perot interferometer and the optical fiber alignment grooves. (c) Optical micrograph of a polymer-coated device.

As expected, in Fig. 5(a), the resonance peak position is found to increase upon exposition to m-xylene, before returning to its initial value when the gas flow is switched back to pure nitrogen. In Fig. 5(b) and (c), this shift of the resonance wavelength is reported, as a function of time, for various concentrations of m-xylene and cyclohexane. The response to 34 ppm m-xylene, the lowest concentration reachable with our flowmeter, is clearly visible. A limit of detection below this value is therefore expected.

The resonance wavelength increase ($\Delta\lambda_{\text{Res}}(t)$) follows an essentially exponential pattern (i.e.: $\Delta\lambda_{\text{Res}}(t) \propto 1 - e^{-t/\tau}$) with a time constant (τ) that differs for each polymer–analyte combination. In PDMS, the response time is found to be almost 4 times faster for cyclohexane ($\tau = 40$ s) than for m-xylene ($\tau = 150$ s). This faster response time however occurs at the expense of an approximately 4 time lower sensitivity to cyclohexane than to m-xylene.

This trade-off between sensitivity and response time is directly related to the partition coefficient (K_{p-a}) of each polymer–analyte combination. Higher K_{p-a} values yield higher absorbed volume

fractions (ϕ_a) and therefore higher sensitivities. However, higher K_{p-a} also yield higher retention times [19] (i.e. lower diffusion constants), which increase the time required for the analyte to diffuse completely inside the 80 μm deep polymer-filled channel.

In Fig. 6, the resonance wavelength shift ($\Delta\lambda_{\text{Res}}$) for the different m-xylene vapor exposures is compared with the analytical model (Eq. (9b)), and with the finite element (Eq. (11)) bonded and sliding models. The expected response due only to refractive index variations is also included to highlight the influence of mechanical deformations on the sensitivity (a control experiment using undeformable mirrors was also carried to confirm the influence of refractive index variations only. See Appendix A3). The absorbed volume fraction of analyte (ϕ_a) is converted (Eq. (3)) to v/v gas concentrations (C_{air}) in Eqs. (9b) and (11), using a $K_{p-a} = 2090$ partition coefficient for m-xylene between PDMS and air [19]. We also use $V_m = 0.123$ L/mol, and the reported [13] refractive indices of PDMS ($n_p = 1.3959$) and xylenes ($n_a = 1.4802$) at $\lambda = 1550$ nm.

Fig. 4. (a) Experimental setup used for vapor phase analyte generation and concentration adjustments. (b) Experimental setup used to evenly deliver diluted analyte over the sensor chip (which will eventually be positioned under the gas cap but is not shown here).

a.

b.

c.

Fig. 5. (a) Measured transmission spectra of a PDMS-coated deformable FPI sensor upon exposure to a 240 ppm m-xylene vapor concentration. The peak resonance wavelength ("X" marked) is measured by fitting a 4th order polynomial on top of the resonance peaks. Its position increases when gas flow is turned on (left) and then returns to its initial position (right) when the flow is switched back to pure nitrogen. (b and c) Resonance wavelength (λ_{Res}) of a PDMS-coated interferometer over time. The "On" and "Off" markers designate the beginning and the end of exposure to the indicated concentrations of m-xylene (b) and cyclohexane (c).

As expected, the experimental results fall between the bonded and sliding models. The silicon-PDMS adhesion force appears to be important enough to yield a higher experimental sensitivity than the sliding model prediction. The sensitivity is, however, lower than the bonded model prediction. This could be an indication that PDMS does not bond perfectly with silicon, but is somewhat free to spatially reorganize itself upon expansion. Growing a thin layer of thermal silicon dioxide, prior to polymer coating, could be a way to increase the polymer-interferometer bonding strength and improve sensitivity. It is also possible that the bonding strength

Fig. 7. Experimental response obtained for the four analyte-polymer combinations of m-xylene (XYL), cyclohexane (CY), PDMS, and the phenyl-doped copolymer (PDMS-PDPS). The linear fits are labelled by their corresponding sensitivities. The error bars are calculated from the repeatability (0.5% of full scale) of the two-channel flowmeter. As cyclohexane is more volatile than m-xylene, low concentration measurements suffer from greater uncertainty (m-xylene error bars are too small to appear on graph).

is already high enough, but that finite element bonded sensitivity was overestimated. As discussed in Section 2.1.2 (see also Appendix A2), small variations of the PDMS Poisson's ratio tend to strongly affect the bonded model sensitivity, which rapidly reaches that of the simplified analytical model.

The simplified analytical model (Section 2.1.1) is found to reproduce our experimental results remarkably well. A relatively good correspondence was expected since, as shown in Eqs. (9b) and (11), the simplified analytical model yields an intermediate sensitivity between the two extreme possible cases (i.e., the bonded and sliding finite element models). Therefore, for other polymers having different mechanical properties, this model could most likely provide a useful estimate of the sensitivity without repeating time consuming finite element simulations. A perfect match with the experimental results, such as in Fig. 6, is however likely to be coincidence and should not be expected in all cases.

As discussed in Section 2.1.1, the simplified analytical model also describes the behavior of out-of-plane deformable FPI sensors [8–11]. Its correspondence with the current experimental results therefore indicates that the in-plane implementation of deformable FPI did not cause any significant sensitivity reduction compared to out-of-plane designs.

In Fig. 7, the experimental response is reported for m-xylene-PDMS (XYL-PDMS) and also for the three other polymer-analyte combinations. The response to both analytes is essentially linear, within the repeatability of the flow meter (0.5% of full scale for each channel). The highest sensitivity (0.023 nm/ppm) is obtained for m-xylene-PDMS-PDPS. This may be due to the addition of phenyl groups within the polymer, which could increase the polymer affinity for aromatic compounds, such as m-xylene. There are, however, other differences between the two polymers, including chain length and the type and degree of cross-linking, so other mechanical and chemical factors may also contribute to the sensitivity difference. The lowest sensitivity (0.0030 nm/ppm) is obtained for cyclohexane (CY) in PDMS, which may indicate that the partition coefficient (K_{p-a}) is significantly lower for cyclohexane than for m-xylene. We did not find measurements of K_{p-a} for CY-PDMS in the literature, but taking the linear temperature-programmed retention index of cyclohexane [20] (LTPRI = 650) and the relations reported in ref. [19], we expect $K_{p-a} \approx 320$ –350. Using these values, the CY-PDMS experimental results are within the bonded and sliding models predictions.

The CY-PDMS-PDPS sensitivity (0.0058 nm/ppm) is almost two times higher than the CY-PDMS response. This was not expected initially since cyclohexane is a non-polar analyte, and since the addition of phenyl groups to PDMS tends to increase its polarity [21]. It is possible that the phenyl groups also increase the Hildebrand solubility parameter of PDMS (initially $\delta = 7.3 \text{ cal}^{1/2} \text{ cm}^{-3/2}$)

Fig. 6. Experimental and simulated (bonded, sliding and simplified analytical models) resonance wavelength shift ($\Delta\lambda_{\text{Res}}$) as a function of m-xylene concentration. "RI only" designates the expected response due to refractive index variations only (i.e., neglecting mechanical deformations).

to a value closer to that of cyclohexane ($\delta = 8.2 \text{ cal}^{1/2} \text{ cm}^{-3/2}$) [1]. This phenomenon was previously reported [21,22] and could explain a higher K_{p-a} for CY-PDMS–PDPS than for CY-PDMS. However, as noted above, there are other differences between the films that may also contribute to the observed K_{p-a} difference, including chain length and degree of cross-linking. Moreover, those differences may also affect the polymer mechanical properties, which could influence the sensitivity.

In Fig. 7, PDMS–PDPS showed a 64% increase in sensitivity to m-xylene and a 93% increase in sensitivity to cyclohexane as compared to PDMS. Having distinct sensitivity differences for each analyte could be useful for the development of artificial olfaction systems. Distinct sensitivity differences were exploited previously for artificial olfaction with other micromechanical sensors, such as cantilevered sensors [23], quartz crystal microbalances (QCM), surface acoustic wave (SAW) sensors [24,25], and out-of-plane deformable FPI sensors [10]. As the sensitivities of our sensors match those of out-of-plane FPIs (see discussion related to Fig. 6), it is likely that exploiting distinct sensitivity differences with our sensors could also allow specific detection.

For micromechanical sensors, the sensor limit of detection (LOD) is usually defined as the analyte concentration that produces a response corresponding to 3 times the standard deviation (std) of the background noise signal [23,26]. We calculated a std = 0.012 nm amplitude noise from the traces of Fig. 5(b and c), and also from the trace of a PDMS–PDPS coated device. Using the sensitivities of the PDMS–PDPS coated devices given in Fig. 7, we therefore expect LODs of 1.6 ppm m-xylene and 6.3 ppm cyclohexane. For comparison, previously reported LODs of QCM [26] and SAW [27] devices to xylenes vapors are on the same order of magnitude, at respectively 1.2 ppm and 3.5 ppm. The reported devices could therefore potentially compete with these technologies, which already find applications in artificial olfaction systems [24,25].

The reported LOD is however more than one order of magnitude larger than what was reported using out-of-plane FPIs [10]. However, as discussed above, the resonance wavelength sensitivity of the reported devices, and of out-of-plane FPIs, are similar (see discussion related to Fig. 6). The significantly better LOD of out-of-plane devices is indeed not achieved through higher device sensitivities, but through optimization of a free-space optical interrogation setup [10] (as opposed, in our case, to passively aligned optical fibers). The reported in-plane devices, although less precise in term of LOD, could therefore still be advantageous in situations that would prevent on-site free-space alignment (e.g. field deployment for industrial artificial olfaction).

4. Conclusions

We reported a chemical sensor based on in-plane silicon Fabry–Perot interferometers functionalized with PDMS-based polymers. Upon analyte absorption, polymer swelling and refractive index variations were shown to induce strong shifts of the interferometer resonance wavelengths. Mechanical deformations were demonstrated to be the dominant sensing mechanism, in accordance with analytical and finite element models. The proposed analytical model is also expected to describe accurately the behavior of out-of-plane FPIs [8–11], and could therefore be useful for sensors relying on such optical configuration.

Fabry–Perot interferometers were functionalized with two different polymers (PDMS and PDMS–PDPS) and were used to detect and quantify two volatile organic compounds (m-xylene and cyclohexane). PDMS–PDPS devices were found to be 64% more sensitive to m-xylene and 93% more sensitive to cyclohexane than PDMS devices. Such distinct sensitivity differences, between sensor coatings, are promising for the development of artificial olfaction

systems. Moreover, the expected limits of detection of the current proof-of-concept devices (1.6 ppm m-xylene, 6.3 ppm cyclohexane) are already on the same order of magnitude as achieved with micromechanical sensing technologies used for artificial olfaction.

Compared with other types of micromechanical sensors, a possible advantage of deformable in-plane FPI sensors is their potential for field deployment and remote interrogation. The proposed sensor does not need any local source of energy to operate. Sensor heads could be deployed in industrial workplaces or for environmental monitoring, and would be linked to an interrogation system only through passively aligned single mode optical fibers. Using a $1 \times N$ optical switch would also allow the readout of multiple sensors (for various locations and/or for various polymers) using a single tunable laser and photodetector.

Compared with previously reported out-of-plane FPIs [8–11], the main distinctive feature of the proposed in-plane sensors is integrability. The in-plane configuration allowed monolithic integration of microfluidic systems, which simplified parallel functionalization of multiple interferometers with different polymers. Integrated fiber alignment grooves also allowed passive alignment of optical fibers, facilitating remote interrogation by avoiding the need for on-site free-space alignment. Finally, successful interfacing of swellable polymers with a silicon optical microsystem is, by itself, an interesting achievement. Such hybrid integration could allow great flexibility in the design of other interferometric, micromechanical, or even electromechanical systems that could be used to maximize sensitivity to polymer swelling.

Acknowledgement

This work was supported by the National Science and Engineering Research Council of Canada (NSERC).

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.snb.2013.02.016>.

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